

## Note

### New approach to the complexation of iron(III) with fructose

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The complexes of iron(III) with monosaccharides have an important role in the transport of this metal across cell membranes<sup>1,2</sup>. The complexation properties of monosaccharides maintain the iron(III) in solution in a low molecular weight, soluble, and stable configuration over a wide pH range<sup>3</sup>. Iron(III)–sugar complexes can be isolated from aqueous solution by the addition of ethanol to a final concentration of 80% (v/v)<sup>3</sup>. The characterization of the isolated complexes of iron(III) with various kinds of sugars and polyols have been performed by molecular weight determination, Mössbauer and infrared spectroscopy<sup>4,5</sup>, EPR spectroscopy<sup>6</sup>, and extended X-ray absorption fine structure spectroscopy (EXAFS)<sup>7</sup>. It has been considered for a long time that the chelating ability of sugars is associated with the dihydroxyacetone structure, which is found in the open-chain fructose molecule<sup>8</sup>. It was proposed that, in these complexes, the iron atom is incorporated into a six-membered ring, bound through the hydroxyl oxygen atoms to C-1 and C-3 of the fructose molecule. Nagy et al.<sup>7</sup> also proposed the bonding of the open-chain form of fructose, but through the hydroxyl-oxygen atoms on C-1 and C-2; this kind of bonding was first proposed by Charley et al.<sup>3</sup>.

On the other hand, the binding of fructose to other elements, such as Ca, Mg,  $\text{UO}_2^{2+}$ , Zn, Cd, Hg, and Al, was found to be in a cyclic form either as a pyranose and/or furanose ring<sup>9–12</sup>.

In an aqueous solution of D-fructose, an equilibrium is established between  $\alpha$ -D-fructopyranose,  $\beta$ -D-fructopyranose,  $\alpha$ -D-fructofuranose,  $\beta$ -D-fructofuranose, and the acyclic isomer. The actual composition depends on the pH, temperature, and other factors. A solution of D-fructose in  $\text{D}_2\text{O}$  at 30°C contains 2%  $\alpha$ -pyranose, 70%  $\beta$ -pyranose, 5%  $\alpha$ -furanose, and 23%  $\beta$ -furanose<sup>13</sup>. The open-chain form was found by  $^{13}\text{C}$  NMR spectroscopy at 80°C only in a small amount (3%)<sup>14</sup>.

Since  $^{13}\text{C}$  NMR spectroscopy is a very useful method for the identification and characterization of carbohydrates and their complexes, in this paper the  $^{13}\text{C}$  NMR spectrum of the Fe–D-fructose complex is reported in order to find out whether iron(III) is the only element to bind an open-chain form of fructose. In the  $^{13}\text{C}$  NMR spectrum of the free ligand, D-fructose, eighteen signals were present,



corresponding to the  $\alpha$ - and  $\beta$ -furanose and  $\beta$ -pyranose forms of D-fructose assigned according to data reported by Angyal and Bethell<sup>15</sup>. The open-chain form was not detected; there was no signal at 214 ppm, which would correspond to the chemical shift of the carbonyl group. In the  $^{13}\text{C}$  NMR spectrum of the Fe–D-fructose complex (Fig. 1), six signals were detected corresponding to the  $\beta$ -pyranose ring form of D-fructose (C-1, 64.85; C-2, 98.91; C-3, 68.43; C-4, 70.52; C-5, 70.06; and C-6, 64.26 ppm). They were almost unchanged in position, but some change in intensity occurred. The chemical shift of the C-1 atom at 64.85 ppm suffered a marked diminution in apparent intensity. The results of  $^{13}\text{C}$  NMR spectroscopy indicate that iron(III) binds preferably to fructose in the  $\beta$ -pyranose ring form which is present in an equilibrium solution of D-fructose in the greatest amount.

## EXPERIMENTAL

D-(–)-fructose (Merck, Germany) and  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$  (Merck, Germany) were reagent grade.

The complex was prepared by dissolution of  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$  (0.02 mol) and D-(–)-fructose (0.2 mol) in  $\text{H}_2\text{O}$  (100 mL). The pH of the solution was adjusted to 11.0 by the addition of 1 M NaOH. A deep-brown solution was obtained. The Fe(III)–D-fructose complex was precipitated by the addition of EtOH and isolated by centrifugation. The precipitate was redissolved in a small volume of water, the pH was adjusted again to 11.0, and the precipitation was repeated. The isolated compound was very soluble in water.

The deduced formula for the Fe–D-fructose complex is  $\text{Fe}_2(\text{C}_6\text{H}_{12}\text{O}_6)_2(\text{OH})_7\text{Na}$ . Anal. Calc.: C, 23.46; H, 5.05; Fe, 18.20; Na, 3.74. Found: C, 23.14; H, 4.43; Fe, 16.67; Na, 3.66.

The  $^{13}\text{C}$  NMR spectra of the solutions of D-fructose (1 M) and the Fe–D-fructose complex (0.2 M) in  $\text{D}_2\text{O}$  were measured on a Varian Gemini 300 Fourier-transform spectrometer at room temperature in 5-mm o.d. tubes at 75 MHz. The spectral width was 19000 Hz; pulse width, 5.0  $\mu\text{s}$ ; pulse width ( $90^\circ$ ), 15.0  $\mu\text{s}$ ; pulse delay, 0.5 s for D-fructose and 2.0 s for Fe–D-fructose. Chemical shifts were measured relative to that of internal 1,4-dioxane, set at 67.4 ppm downfield from that of  $\text{Me}_4\text{Si}$ .

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## REFERENCES

- 1 P. Saltman, *J. Chem. Educ.*, 42 (1965) 682–687.
- 2 Th.G. Spiro and P. Saltman, *Struct. Bonding*, 6 (1969) 117–156.

- 3 P.J. Charley, B. Sarkar, C.F. Stitt, and P. Saltman, *Biochim. Biophys. Acta*, 69 (1963) 313–321.
- 4 M. Tonković, S. Musić, O. Hadžija, I. Nagy-Czako, and A. Vertes, *Acta Chim. Acad. Sci. Hung.*, 110 (1982) 197–202.
- 5 M. Tonković, O. Hadžija, and I. Nagy-Czako, *Inorg. Chim. Acta*, 80 (1983) 251–254.
- 6 L. Nagy, K. Burger, J. Kuerti, M.A. Mostafa, L. Korecz, and I. Kiricsi, *Inorg. Chim. Acta*, 124 (1986) 55–59.
- 7 L. Nagy, H. Ohtaki, T. Yamaguchi, and M. Nomura, *Inorg. Chim. Acta*, 159 (1989) 201–207.
- 8 P.S. Davis and D.J. Deller, *Nature (London)*, 212 (1966) 404–405.
- 9 W.J. Cook and C.E. Bugg, *Acta Crystallogr., Sect. B*, 32 (1976) 656–659.
- 10 H.A. Tajmir-Riahi, *Biophys. Chem.*, 23 (1986) 223–228.
- 11 H.A. Tajmir-Riahi, *Carbohydr. Res.*, 172 (1988) 1–10.
- 12 M. Tonković, H. Bilinski, and M.E. Smith, *Inorg. Chim. Acta*, 197 (1992) 59–65.
- 13 S.J. Angyal, *Adv. Carbohydr. Chem. Biochem.*, 42 (1984) 15–68.
- 14 W. Funcke, C. von Sonntag, and C. Triantaphylides, *Carbohydr. Res.*, 75 (1979) 305–309.
- 15 S.J. Angyal and G.S. Bethell, *Aust. J. Chem.*, 29 (1976) 1249–1265.